

## BBA Report

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### EFFECT OF ANIONS ON POTASSIUM SELF-EXCHANGE IN ASCITES TUMOR CELLS

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#### Summary

Potassium self-exchange in ascites cells was partially suppressed when chloride in the incubation medium was replaced by bromide, and completely abolished in iodide-, nitrate- or sulfate-containing saline. Thus, it appears that potassium self-exchange activity is anion-dependent.

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One of the modes by which monovalent cations are transferred across cell membranes is self-exchange [1]. In this communication we present evidence that  $K^+$  self-exchange in ascites cells is dependent on the anion composition of the medium.

Fig. 1A shows the effect of ouabain and furosemide on the influx of  $^{86}Rb$  into ascites cells; this isotope serves as a tracer for  $K^+$  in this system (e.g., ref. 2). In the absence of furosemide ouabain had a small inhibitory effect. Furosemide alone decreased the rate of  $^{86}Rb$  influx by 50%. Both inhibitors together brought about a marked reduction to approximately 8% of the control value. These findings are similar to those of Tupper [2]. Ouabain caused a net loss of  $K^+$  (Table I); we show elsewhere (unpublished) that the concentration of ouabain employed is sufficient to totally inhibit the  $(Na^+ + K^+)$  ATPase. Furosemide did not influence the cellular  $K^+$  content (Table I). In another experiment (Table II) the furosemide-sensitive component of  $^{86}Rb$  influx was shown to be equal in absolute magnitude to that of efflux, both in the absence and in the presence of ouabain; in the presence of ouabain the furosemide-sensitive component was markedly stimulated, as in the experiment of Fig. 1.

Fig. 1B shows the results of a similar experiment carried out in a saline solution in which 100 mM  $NaNO_3$  was substituted for  $NaCl$ . In this system furosemide had no effect. Moreover, all fluxes were reduced to the rates observed in the presence of furosemide in  $Cl^-$  saline (1.9 and  $0.3 \mu mol K^+ \cdot g^{-1}$

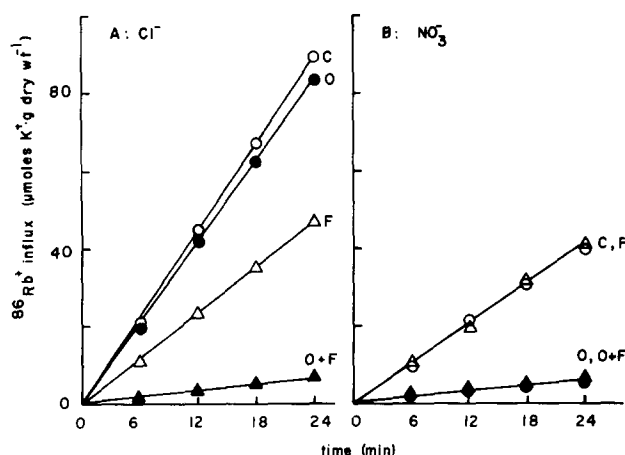


Fig. 1. Effect of inhibitors on  $^{86}\text{Rb}$  influx into ascites cells in  $\text{Cl}^-$ -saline and  $\text{NO}_3^-$ -saline. Ascites cells were grown in the abdominal cavities of Swiss Webster mice, and harvested after 6–12 days. They were washed in a saline solution containing: 50 mM *N*-2-hydroxyethylpiperazine-*N*'-2-ethanesulfonic acid (HEPES) anion, neutralized with 20 mM NaOH and 4.4 mM KOH, 1 mM  $\text{CaCl}_2$ , 1 mM  $\text{MgSO}_4$ , 20 mM glucose, and either 100 mM NaCl or 100 mM  $\text{NaNO}_3$ . The final pH was 7.5. They were then suspended at a concentration of 1.5% in the respective saline plus or minus furosemide (1.2 mM), and incubated for 90 min at  $37^\circ\text{C}$  to allow the ion content to stabilize. At time zero  $^{86}\text{Rb}$  (20 nCi/ml) was added alone or together with ouabain (1.2 mM). At the indicated times, duplicate 3 ml samples of suspension were spun down at 1500 rev./min, the supernatant decanted and saved, and the cells extracted with 5% trichloroacetic acid. Radioactivity of cell extracts and supernatants was counted by liquid scintillation counting, and  $\text{K}^+$  content determined by flame photometry (see Table I). Extracellular space correction was made by extrapolating the uptake curves to zero time. Points shown are average of duplicates; duplicates were equal within 5%. A,  $\text{Cl}^-$ -saline; B,  $\text{NO}_3^-$ -saline.  $\circ$  (C), control;  $\bullet$  (O), ouabain;  $\triangle$  (F), furosemide;  $\blacktriangle$  (O + F), ouabain plus furosemide.

dry wt.  $\cdot \text{min}^{-1}$ , in the absence and presence of ouabain, respectively).  $\text{K}^+$  contents were comparable to those in the  $\text{Cl}^-$  system (Table I).

From these data it appeared that either removal of  $\text{Cl}^-$  or addition of  $\text{NO}_3^-$  abolished the furosemide-sensitive components of  $^{86}\text{Rb}$  influx. In order to further examine this point we investigated the effect of several other anions in the presence of ouabain, where the size of the furosemide-inhibitable component was larger. Fig. 2 shows the results of these experiments. The dotted line gives the level of influx in the presence of furosemide and ouabain, i.e. the non-mediated influx. This level was the same in all saline systems tested ( $0.37 \pm 0.02 \mu\text{mol K}^+ \cdot \text{g}^{-1} \text{ dry wt.} \cdot \text{min}^{-1}$ ). As  $\text{Cl}^-$  was gradually replaced by  $\text{I}^-$ ,  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$ , the furosemide-sensitive component of the flux was reduced; at 2 mM  $\text{Cl}^-$  it was completely abolished. In contrast,  $\text{Br}^-$  at 90 mM suppressed

TABLE I  
EFFECT OF INHIBITORS AND NITRATE ON  $\text{K}^+$  CONTENT

Samples were obtained from the experiment shown in Fig. 1, 24 min after addition of  $^{86}\text{Rb}$  plus or minus ouabain. Values given are the average of duplicates; duplicates were equal within 5%.

Major anion	$\text{K}^+$ content ( $\text{mmol} \cdot \text{g}^{-1}$ dry wt.)			
	$\text{Cl}^-$		$\text{NO}_3^-$	
	- ouabain	+ ouabain	- ouabain	+ ouabain
- Furosemide	0.31	0.25	0.32	0.27
+ Furosemide	0.30	0.26	0.32	0.27

the furosemide-sensitive component by only 35%. Cellular  $K^+$  content and the net loss induced by ouabain (not shown) were equal for all anions tested except  $SO_4^{2-}$ , and comparable to those in Table I. In  $SO_4^{2-}$  saline, values were approximately 24% lower on a dry weight-basis; this is probably due to shrinkage of the cells [3].

Tupper [2] interpreted his data on the effect of furosemide to mean that this compound inhibited  $K^+$  self-exchange. For the experiments presented here this interpretation is supported by the observation that the furosemide-inhibitable fractions of influx and efflux were equal in absolute magnitude (Table II), as well as the fact that furosemide did not alter  $K^+$  content (Table I). It thus seems justified to interpret the furosemide-inhibitable  $^{86}Rb$  flux components as arising from  $K^+$  self-exchange. Our data show that this self-exchange is influenced by the anionic composition of the medium. The concentration dependence of this effect was the same for  $I^-$ ,  $NO_3^-$ , and  $SO_4^{2-}$  (Fig. 2). This suggests that  $Cl^-$  is required for  $K^+$  self-exchange with a rather high apparent  $K_m$ .  $Br^-$  appears partially able to substitute for  $Cl^-$  in sustaining  $K^+$  self-exchange. Effects of anions on unidirectional cation fluxes comparable to those described here have been reported by Funder and Wieth [4] for human red blood cells.

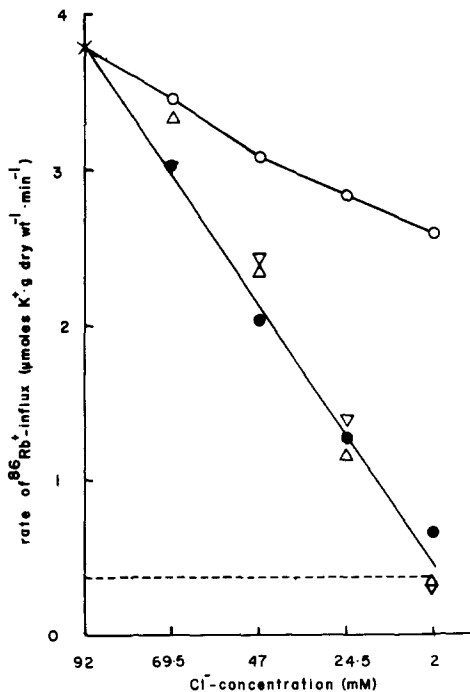


Fig. 2. Effects of different anions on the ouabain-inhibited influx of  $^{86}Rb$ . Saline solutions were similar to that described in the legend to Fig. 1, with either  $NaBr$ ,  $NaI$ ,  $NaNO_3$  (100 mM) or  $Na_2SO_4$  (50 mM) substituted for  $NaCl$ .  $Cl^-$ -saline was mixed with the other salines to give the indicated  $Cl^-$  concentrations. The experiment was performed as described for Fig. 1. Rates were calculated from the difference in cellular radioactivity between 30 min and 10 min after the addition of  $^{86}Rb$  (20 nCi/ml) plus ouabain (1.2 mM). The dotted line gives the influx rate for the five unmixed saline solutions in the presence of furosemide (1.2 mM).  $I^-$ , which was found to be a strong quencher in our scintillation system, was precipitated with  $AgNO_3$  prior to counting of radioactivity.  $\circ$ ,  $Br^-$ ;  $\bullet$ ,  $I^-$ ;  $\Delta$ ,  $NO_3^-$ ;  $\nabla$ ,  $SO_4^{2-}$ .

TABLE II

EFFECT OF INHIBITORS ON  $^{86}\text{Rb}$  INFLUX AND EFFLUX IN  $\text{Cl}^-$  SALINE

For the rate of  $^{86}\text{Rb}$  efflux, ascites cells were equilibrated for 90 min in the presence of the isotope (10 nCi/ml). At the end of this period cells were spun down for 1 min at 1000 rev./min, and at time zero the pellet was resuspended at a concentration of 1.5% in  $\text{Cl}^-$  saline in the presence or absence of inhibitors (concentration, 1.2 mM). Radioactivity remaining in the cells at a given time was determined as described in the legend to Fig. 1. Efflux rate constants ( $k_e$ ) were obtained by linear regression from semilogarithmic plots of the decrease in cellular radioactivity with time. Correlation coefficients ( $r$ ) exceeded 0.94 in all cases. Total efflux was calculated from the product of  $k_e$  and the average of the cellular  $\text{K}^+$  contents at zero time and 24 min. Influx was determined in the same experiment by methods described in the legend to Fig. 1; the external  $\text{K}^+$  concentration was 2.5 mM. The values for the furosemide-sensitive components were obtained by subtraction from the values given for the unidirectional fluxes; the error in these derived values can be estimated at about 10%.

	Efflux rate constant $k_e$ ( $\text{min}^{-1}$ )	Unidirectional fluxes ( $\mu\text{mol K}^+ \cdot \text{g}^{-1} \text{ dry wt.} \cdot \text{min}^{-1}$ )		Furosemide-sensitive components ( $\mu\text{mol K}^+ \cdot \text{g}^{-1} \text{ dry wt.} \cdot \text{min}^{-1}$ )	
		Influx	Efflux	Influx	Efflux
—	0.018	7.1	6.8		
+ Furosemide	0.007	3.2	2.8	3.9	4.0
+ Ouabain	0.034	7.1	10.8	6.8	8.2
+ Furosemide + ouabain	0.008	0.3	2.6		

The nature of the anion effect on  $\text{K}^+$  self-exchange is not clear. Apparently the passive permeability of the anion is not involved since  $\text{Cl}^-$  and  $\text{NO}_3^-$  have comparable passive permeabilities (unpublished data), whereas that of  $\text{SO}_4^{2-}$  has been reported to be considerably lower [3]. A clue to the mechanism of  $\text{K}^+$  self-exchange and its susceptibility to the anion composition of the medium may possibly be found in the observation of Heinz et al. [5] that  $\text{K}^+$  and  $\text{Cl}^-$  are co-transported under certain conditions.

The fact that  $\text{K}^+$  self-exchange was stimulated in the presence of ouabain suggests a close relation between pumping and exchange activities. This relation (often of a complementary nature) has been observed in a number of systems [6].

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## References

- 1 Glynn, I.M. and Karlish, S.J.D. (1975) *Annu. Rev. Physiol.* 37, 13–55
- 2 Tupper, J.T. (1975) *Biochim. Biophys. Acta* 394, 586–596
- 3 Levinson, C. and Villeral, M.L. (1974) *J. Cell. Physiol.* 85, 1–14
- 4 Funder, J. and Wieth, J.O. (1967) *Acta Physiol. Scand.* 71, 168–185
- 5 Heinz, E., Geck, P., Pietrzyk, C., Burckhardt, G. and Pfeiffer, B. (1977) *J. Supramol. Struct.* 6, 125–133
- 6 Mills, B. and Tupper, J.T. (1976) *J. Cell. Physiol.* 89, 123–132